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ORIGINAL ARTICLE

Two new alkaloids from *Gelsemium elegans*

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Two new alkaloids, named gelsenine (1) and 11-methoxyhumantenmine (2), were isolated from the whole plant of *Gelsemium elegans*. The structures were elucidated on the basis of 1D NMR, 2D NMR, and MS methods.

Keywords: Gelsemium elegans; gelsenine; 11-methoxyhumantenmine; oxindol alkaloid

1. Introduction

Gelsemium elegans Benth. is a well-known toxic plant in southeastern Asia, which is used in Chinese folk medicine as an analgesic, antispasmodic, and antitumor agent [1]. The continuing interest in the chemistry of Gelsemium alkaloids comes largely from the potent biological activity and special chemical structure [2-10]. As part of our systematic studies on the chemical constituents of Chinese medicinal plants, we initiated a chemical study on G. elegans. By the general isolation methods, two new alkaloids, named gelsenine (1) and 11-methoxyhumantenmine (2) (Figure 1), were isolated from the whole plant. On the basis of a spectroscopic analysis, these chemical structures were determined. In this paper, the isolation and structural elucidation of these compounds are described.

2. Results and discussion

Compound 1 was obtained as a pale yellow amorphous powder. The molecular

formula $C_{20}H_{26}N_2O_4$ was confirmed by HR-MS at m/z 359.1977 [M+H]⁺. It gave a brown spot in normal-phase TLC (CHCl₃-MeOH, 90:10) with Dragendorff-Wagner (1:1) reagent. The UV absorption maxima of **1** at 209 (log ε 4.25) and 257 nm (log ε 3.80) revealed an oxindole chromophore [3]. In the IR spectrum, **1** showed absorption bands at 3423 cm⁻¹ (NH group), 1705 cm⁻¹ (carbonyl group), 1616 and 1463 cm⁻¹ (aromatic ring).

The ¹H NMR spectrum of **1** showed four aromatic protons due to the ring of the oxindole system, a N_a -methoxy group at δ 4.02 (3H, s), a methine group bearing amine nitrogen at δ 3.78 (br s, H-5), an oxymethine proton at δ 3.55 (d, H-3), oxymethylene protons at δ 4.27 (br s, H-18), oxymethylene protons at δ 3.28 (d) and 3.47 (d, H₂-17), and an ethyl group at δ 0.91 (3H, t, H₃-20) and 1.95 (2H, q, H₂-21). The ¹³C NMR spectrum of **1** showed 20 carbons, including a methine

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Figure 1. Chemical structures of compounds 1 and 2.

bearing a nitrogen atom at δ 59.3 (C-5), a quaternary carbon bearing a nitrogen atom at δ 69.8 (C-16), an oxygenated methine at δ 74.8 (C-3), and two oxygenated methylenes at δ 62.7 (C-17) and 63.4 (C-18).

In the HMBC spectrum, correlations of H-3 with C-6, C-8, and C-14, H-6 with C-2, C-8, C-3, and C-7, H-18 with C-5, C-14, and C-19, H-15 with C-3 and C-16, H-14 with C-5 and C-15, H-17 with C-14, C-15, and C-21, H-21 with C-16 and C-17 allowed to deduce the presence of three six-member rings which are connected to each other (Figure 2).

The stereochemistry of **1** was determined by the NOE correlations (Table 1). H-18 showed a clear correlation with H-9, suggesting that this compound has a C-19a-H. This was further confirmed by the NOESY correlation between H-19 and H-15a. The NOE correlation between H-9 and one of the C-6 methylene proton signals at δ 2.18 led to the assignment



Figure 2. Selected HMBC correlations of compound 1 ($H \rightarrow C$).

of this signal as H-6, whereas the remaining signal at δ 2.04 is for H-6 β . The NOE correlation between H-20, H-21, and H-17 β indicates the probable assignment of the proton NMR resonances of the C-17 methylene protons. The NOE correlation between H-15a and H-19 also indicates the assignment of the NMR proton resonances of C-15 methylene protons. Thus, all of the protons and functional groups were assigned and the complete structure of 1 was elucidated, named gelsenine. The HMBC, ¹H-¹H COSY, and NOESY results of 1 led to assignments for the ¹H and ¹³C signals, as shown in Table 1.

Compound 2 was obtained as a pale yellow amorphous powder. The molecular formula C₂₀H₂₄N₂O₄ was confirmed by HR-MS at m/z 357.1815 [MM+H]⁺. It gave a brown spot in normal-phase TLC (CHCl₃-MeOH, 90:10) with Dragendorff-Wagner (1:1) reagent. The UV absorption maxima at 257 and 209 nm revealed an oxindole system [3]. The ¹H NMR spectrum displayed three aromatic proton signals at δ 7.41 (1H, d, J = 8.3 Hz, H-9), 6.56 (1H, dd, J = 8.3, 2.4 Hz, H-10), and 6.47 (1H, d, J = 2.4 Hz, H-12), forming an ABX system; two methoxyl signals at δ 3.94 (3H, s) and 3.81 (3H, s); one ethyl signal at δ 1.29 (3H, t, $J = 7.4 \,\mathrm{Hz}, \,\mathrm{H-18}, \,2.73 \,(1\mathrm{H},$ dq, $J = 17.0, 7.4 \text{ Hz}, \text{ H-19}\alpha$), and 2.45 (1H, dq, J = 17.0, 7.4 Hz, H-19 β). The ¹³C NMR and HMQC spectra of compound 2 exhibited 20 carbon signals, including one

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15α, 15β, 17β 15α, 15β, 17β 12 NOESY^c 18 19, 20, 21 20, 21 $6\alpha, 9, 14$ 3, 6α, 18 OCH₃ 20,21 9.18 15α 6 5, 14, 18 21 20 6a, 19 5, 6β 6a 9, 11 10, 12 10 15β l7β 17a 19 4 Ξ 3, 158, 19 14, 15a COSY Three-bond 2, 3, 8 3 15, 21 5, 14 8, 12 9 15, 21 6, 8, 17 7, 11, 13 10 5 3 110 110တ် HMBC^b Two-bond 15 14, 16 14, 16 12 9 10 21 16, 20 4 $\neg \neg$ 16 19 36.2 22.9 74.8 59.3 125.3 123.9 128.4 107.3 137.9 69.8 23.0 174.7 131.3 62.7 39.5 9.4 63.4 33.4 57.3 63.4 ŝ 2.18 (1H, dd, *J* = 16.0, 3.4 Hz) 2.04 (1H, d, *J* = 16.0 Hz) 2.04 (1H, m) 2.28 (1H, d, *J* = 14.0 Hz) 2.11 (1H, over) 3.28 (1H, d, J = 10.4 Hz)3.47 (1H, d, J = 10.4 Hz)3.55 (1H, d, J = 6.0 Hz)7.40 (1H, d, J = 7.6 Hz)6.97 (1H, d, J = 7.6 Hz)1.95 (2H, q, J = 7.4 Hz)7.14 (1H, t, J = 7.6 Hz)7.31 (1H, t, J = 7.6 Hz) 0.91 (3H, t, J = 7.4 Hz) $\delta_{\rm H}$ 2.79 (1H, br m) 3.78 (1H, br s) 4.27 (2H, br s) 4.02 (3H, s) -N-0-CH₃ Position 6] 20 5

Table 1. ¹H NMR (300 MHz) and 13 C NMR (75 MHz) spectral data of gelsenine (1).^a

^b Numbers in each column, respectively, indicate the carbons coupled with the proton through two or three bonds. Notes: ^aCDCl₃ was used as a solvent; chemical shifts (δ) in ppm; coupling constants J in Hz. ° NOESY spectra were obtained at 600 MHz.

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Table 2. ¹H NMR (400 MHz) and ¹³C NMR (100 MHz) spectral data of compound 2^{a} .

Position	$\delta_{\rm H}$ (Hz)	$\delta_{\rm C}$
2		171 7
3	3 71 (dd 4 5 2 0)	75.3
5	4.40 (m)	72.1
5	2.27 (m)	27.9
0a 60	2.37 (III) 2.27 (dd $14.2, 2.2$)	57.0
ор 7	2.27 (du, 14.3, 2.3)	55 A
/		33.4
8		124.0
9	7.41 (d, 8.3)	125.4
10	6.56 (dd, 8.3, 2.4)	107.8
11		160.1
12	6.47 (d, 2.4)	93.9
13		139.1
14a	2.36 (m)	26.9
14β	2.14 (ddd, 14.8, 10.0,	
	4.7)	
15	2.87 (t, 9.2)	42.5
16	2.57 (t, 8.1)	39.7
17	4.27 (2H, m)	62.0
18	1.29 (3H. t. 7.4)	9.9
19a	2.73 (dg 17.0.7.4)	25.6
19B	2.45 (dq, 17.0, 7.4)	20.0
20	2.15 (aq, 17.6, 7.1)	184.8
	3 81 (3H s)	55.5
	$2.04(2H_{c})$	55.5 62.4
IV UCH3	5.74 (50, 8)	05.4

Note: ^aCDCl₃ was used as a solvent. Chemical shifts (δ) in ppm.

carbonyl carbon, six aromatic carbons, two methoxyl carbons, one methyl carbon, four methylene carbons, four methine carbons, one quaternary carbon, and one imine carbon. The ¹H NMR and ¹³C NMR spectra (see Table 2) were very similar to humantenmine [11] except for the signals in the aromatic region, namely the lack of an aromatic proton signal and the existence of a methoxyl group at δ_H 3.81 and δ_C



Figure 3. Selected HMBC correlations of compound 2 ($H \rightarrow C$).

55.5. The HMBC correlations (Figure 3) between the proton at δ 3.81 (3H, s) and the carbon at $\delta_{\rm C}$ 160.1 indicated that a methoxyl group was attached to the oxindole ring at C-11. Furthermore, the HMBC correlations between the protons at δ 2.73 (H-19 α), 2.45 (H-19 β) and the carbons at $\delta_{\rm C}$ 184.8 (C-20), 42.5 (C-15) showed that an ethyl group was attached to the imine group C-20. From the above data, compound **2** was deduced to be 11-methoxyhumantenmine.

3. Experimental

3.1 General experimental procedures

UV spectra were recorded on a Shimadzu UV-2501PC spectrometer. Optical rotations were measured on a Perkin-Elmer digital polarimeter. IR spectra were taken on a Perkin-Elmer RX1 FT-IR spectrometer. NMR spectra were recorded in CDCl₃ using TMS as the internal standard on a Bruker-APX-300, Bruker-APX-400, and Bruker-APX-600 instrument. ESI-MS were obtained on a Bruker Esquire 2000 instrument. HPLC was performed on a JASCO LC-2000 instrument with a HiQ sil $C_{18}V$ (250 × 10 mm, 5 µm) column using a UV detector. Column chromatography was carried out on a glass open column, and Sephadex LH-20 (Amersham Biosciences, Shanghai, China) and silica gel (200-300 mesh; Qingdao Haiyang Chemical Co., Ltd, China) were used as adsorbents.

3.2 Plant material

The specimens of *G. elegans* Benth. were collected from the Fujian Province of China (January 2003), and identified by Prof. Qishi Sun, Shenyang Pharmaceutical University, China. Herbarium voucher specimens (GE-03002) are deposited at the Department of Pharmacy, Shenyang Northern Hospital.

3.3 Extraction and isolation

The powdered whole plant of G. elegans was alkalified with 3% NaOH, dried at room temperature, and then extracted with CHCl₃. The chloroform extract was dissolved in the acid water, partitioned with CHCl₃, and then the water layer was alkalified with saturated NaOH, and extracted with CHCl₃ again, to obtain the alkaloids of G. elegans. The alkaloids were further subjected to a silica gel column, eluted with a step gradient solvent system of CHCl₃-MeOH (99:1 \rightarrow 9:1, v/v) to obtain nine fractions (1-9). Fraction 8 (0.73 g) was repeatedly chromatographed on HPLC $(250 \times 10 \text{ mm}, 5 \mu \text{m})$ and eluted with MeOH-H₂O-diethylamine (65:35:0.01, v/v/v) to yield gelsenine (1, 4.1 mg) and 11-methoxyhumantenmine (2, 10.3 mg).

3.3.1 Compound 1

A pale yellow amorphous powder, mp 180–182°C, $[\alpha]_D - 114$ (c = 0.81, MeOH), UV (MeOH) λ_{max} (log ε): 208 (3.90), 254 (3.22) nm. IR (KBr) ν_{max} (cm⁻¹): 3422, 1718, 1615, 1463. ¹H and ¹³C NMR spectral data (see Table 1). HR-FAB-MS: m/z 359.1977 [MM+H]⁺ (calcd for C₂₀H₂₇N₂O₄, 359.1971).

3.3.2 Compound 2

A pale yellow amorphous powder, mp $160-162^{\circ}$ C, $[\alpha]_{\rm D} - 74$ (c = 0.93, MeOH), UV (MeOH) $\lambda_{\rm max}$ (log ε): 209 (4.25), 257 (3.80) nm. IR (KBr) $\nu_{\rm max}$ (cm⁻¹): 3430, 1726, 1629, 1496. ¹H and ¹³C NMR spectral data (see Table 2).

HR-FAB-MS: m/z 357.1815 [MM+H]⁺ (calcd for C₂₀H₂₅N₂O₄, 357.1814).

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